**Multi-component assembly of the bicyclic core associated with the tRNA synthetase inhibitors SB-203207 and SB-203208. Application to the synthesis of biologically active analogues†**

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**The ketone (±)-5, which embodies the bicyclic core associated with the title tRNA synthetase inhibitors 1 and 2, has been prepared** *via* **a three-component coupling reaction involving 2-(hydroxymethyl)cyclopent-2-enone (15), methylamine (6) and propiolamide (10); straightforward elaboration of the readily derived acetates**  $(-)$ **-21 and**  $(+)$ **-21 has provided the biologically active analogues 23 and 24, respectively, of the title compounds.**

The emergence of 'superbugs' such as vancomycin-resistant *Staphylococcus aureus* has prompted extensive efforts to identify new anti-infective agents.<sup>1</sup> High throughput screening regimes have led to the discovery of a number of novel leads including SB-203207 (**1**) and SB-203208 (**2**) which are potent inhibitors of both bacterial and mammalian isoleucyl tRNA synthetases.2 The structurally related natural product altemicidin (3),<sup>3</sup> a novel acaricidal and anti-tumour agent, has been the subject of an elegant total synthesis.<sup>4</sup> However, the methods<sup>4,5</sup> currently available for construction of the hexahydroazaindene core associated with such compounds are unlikely to be practical in providing a broad range of analogues of **1** and **2** for testing as anti-infective agents. On this basis we now describe a multi-component and potentially highly flexible method for construction of the azabicyclic ketones (±)-**4** and (±)-**5** as well as conversion of the latter into biologically active analogues of the title compounds.



In our initial approach to (±)-**4** and (±)-**5** we envisaged that these might be constructed in a one-pot process from methylamine (**6**), formaldehyde (**7**), cyclopent-2-enone (**8**) and the appropriate propiolic acid derivative **9** or **10** (Fig. 1). In particular, it seemed possible that in the presence of a suitable catalyst the Schiff-base (imine) derived from condensation of **6** and **7** could participate in an 'aza-Baylis–Hillman' reaction6 with **8** to give *N*-methyl-2-(aminomethyl)cyclopent-2-enone



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which would then react, through nitrogen in a hetero-Michaeladdition reaction, with **9** or **10**. The enamine–cyclopentenone conjugate thus formed might then be expected to undergo an intra-molecular Michael-addition reaction,<sup>7</sup> thereby providing the target ketones  $(\pm)$ -4 and  $(\pm)$ -5. In the event, mixing the four components **6**–**9** with DABCO, a proven catalyst for the Baylis– Hillman reaction, in water at room temperature (**CAUTION** highly exothermic!) resulted in a complex mixture of products from which the 1,5-diazacycloocta-2,6-diene **13** could be isolated and the structure of which follows from spectroscopic analysis. Clearly, **6**, **7** and **9** but not **8** have been incorporated into this product and further studies revealed that simply mixing the former compounds in water (Scheme 1) provided diene **13** in 45% yield. Presumably, a key intermediate in this conversion



**Scheme 1** *Conditions*: (i) H2O, DABCO (cat.), *ca*. 18 °C, 16 h.

<sup>†</sup> Electronic supplementary information (ESI) available: spectral data for **5**, crystal data for  $(\pm)$ -21 (CCDC 165269), HPLC for  $(+)$ - and  $(-)$ -21. See http://www.rsc.org/suppdata/cc/b1/b104890m/



**Scheme 2** *Reagents and conditions*: (i) DABCO (*ca*. 0.25 mol% wrt **8**), aq. HCHO (1.5 mole equiv.), THF, 18 °C, 23 h; (ii) Ac<sub>2</sub>O (2 mole equiv.), Et<sub>3</sub>N (1.65) mole equiv.), DMAP (cat.), CH2Cl2; (iii) **6** (1.5 mole equiv.), **9** (1.5 mole equiv.), DABCO (1.25 mole equiv.), H2O, 18 °C, 5–7 days; (iv) **14** (1.6 mole equiv.), DABCO (1 mole equiv.), EtOH, 18 °C, 15 h; (v) 14 (1.7 mole equiv.), EtOH, 18 °C, 15 h; (vi) 11 (1 mole equiv.), Pd(PPh<sub>3</sub>)<sub>4</sub> (10 mol%), THF, 18 °C, 10–14 days; (vii) L-Selectride® (1.0 mole equiv. of a 1 M solution in THF), THF,  $-17$  °C, 0.5 h; (viii) 22 (2 mole equiv), Et<sub>3</sub>N (2 mole equiv.), ClCOCOCl (2 mole equiv.), 0 °C, 0.5 h then (+)- or (-)- 20, Et<sub>3</sub>N (1 mole equiv.), DMAP (cat.), DMF, 0 to 18 °C, 1.5 h; (ix) H<sub>2</sub> (1 atm), 10% Pd on C (cat.), MeOH, 18 °C, 4 h.

is the enamine **11**8,9 (resulting from Michael addition of methylamine to methyl propiolate) which condenses with **7** to give the 1-aza-3-methoxycarbonylbuta-1,3-diene **12** that, in turn, undergoes cyclodimerisation to the observed product. An analogous sequence starting with amide **10**, and which would have been presumed to involve intermediate **14**,8 failed to deliver the bis(carboxamide) analogue of compound **13**.

The above-mentioned and ready condensation of **7** with **11**, rather than its participation in an initial Baylis–Hillman reaction with **8**, clearly thwarted attempts to implement the proposed four-component coupling approach to targets (±)-**4** and (±)-**5**. To circumvent such problems, **7** and **8** were subject to a dedicated Baylis–Hillman reaction then an aqueous solution of the resulting 2-(hydroxymethyl)cyclopent-2-enone (**15**)10 (Scheme 2) was treated with **6** and **9** in the presence of stoichiometric amounts of DABCO. In this manner the unstable ketone (±)-**4** was eventually obtained (*ca*. 20% after *ca*. 5 days). An analogous reaction using propiolamide **10** afforded the more stable congener (±)-**5** (*ca*. 20%). A superior method (40% yield after *ca*. 15 h) for producing (±)-**5** involved treating an ethanolic solution of the acetate **16**, derived from alcohol **15**, with **14**<sup>8</sup> (resulting from Michael addition of methylamine to propiolamide) in the presence of DABCO. Surprisingly, the same reaction when carried out in the absence of DABCO afforded the isomeric hexahydroazaindene (±)-**17** (40%) as the major product of reaction. Similarly, when a THF solution of **16** was treated with 11 in the presence of  $(Ph_3P)_4P$ d the structurally related ester (±)-**18** (*ca*. 20%) was obtained.

Diastereofacially selective reduction of ketone (±)-**5** with L-Selectride® yielded the alcohol (±)-**20** (96%), the readily available acetate derivative, (±)-**21** (63%), of which proved suitable for single-crystal X-ray analysis. Alcohol (±)-**20** was readily coupled with the acid chloride derived from **22** and the resulting diastereomeric mixture of esters was subjected to hydrogenolytic deprotection to produce an inseparable and *ca*. 1+1 mixture of **23** and **24**. In an effort to obtain diastereomerically pure samples of these materials several methods for preparing the monochiral forms of ketone **5** were examined but none of the several chiral catalysts that have been used to effect asymmetric Baylis–Hillman reactions11 proved effective in promoting the enantioselective coupling of **14** and **15**. While various chiral ester derivatives of **15** participated in reaction with **14** to produce ketone **5** in acceptable chemical yield, the observed diastereomeric excesses were disappointing  $\left($  < 17%). As a consequence, the racemic acetate (±)-**21** was resolved using chiral HPLC techniques (see ESI†). Coupling of each of the enantiopure alcohols with the acid chloride derivative of **22**

gave, after hydrogenolytic deprotection, the target molecules **23**  $[from (-)-21]$  and 24  $[from (+)-21]$ . Independent testing of 23 and **24** as inhibitors of *S. aureus*-derived IRS12 revealed that the former compound shows an  $IC_{50}$  of 3.7  $\mu$ M while the analogous value for the 'unnatural' diastereoisomer 24 is 12.4  $\mu$ M. Interestingly, this difference in activity is even more pronounced with *S. aureus*-derived LRS (0.42  $\mu$ M *vs.* no inhibition at 100 mM), *S. aureus*-derived VRS (6.35 mM *vs*. no inhibition at 100  $\mu$ M) and rat liver IRS (0.57  $\mu$ M *vs*. 13.5  $\mu$ M).

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## **Notes and references**

- 1 T. F. Gale, J. Görlitzer, S. W. O'Brien and D. H. Williams, *J. Chem. Soc., Perkin Trans. 1*, 1999, 2267. For an up-to-date overview of antibiotic resistance see: C. M. Henry, *Chem. Eng. News*, 2000, **78**, 41.
- 2 A. L. Stefanska, R. Cassels, S. J. Ready and S. R. Warr, *J. Antibiot.*, 2000, **53**, 357; C. S. V. Houge-Frydrych, M. L. Gilpin, P. W. Skett and J. W. Tyler, *J. Antibiot.*, 2000, **53**, 364. For the production and biological evaluation of semi-synthetic analogues of **1** and **2** see: M. G. Banwell, C. F. Crasto, C. J. Easton, A. K. Forrest, T. Karoli, D. R. March, L. Mensah, M. R. Nairn, P. J. O'Hanlon, M. D. Oldham and W. Yue, *Biorg. Med. Chem. Lett.*, 2000, **10**, 2263.
- 3 A. Takahashi, H. Naganawa, D. Ikeda and Y. Okami, *Tetrahedron*, 1991, **47**, 3621 and references cited therein.
- 4 A. S. Kende, *Pure Appl. Chem.*, 1997, **69**, 407 and references cited therein.
- 5 T. Sano, Y. Horiguchi, K. Imafuku and Y. Tsuda, *Chem. Pharm. Bull.*, 1990, **38**, 366; E. W. Baxter, D. Labaree, S. Chao and P. S. Mariano, *J. Org. Chem.*, 1989, **54**, 2893.
- 6 A. Kamimura, Y. Gunjigake, H. Mitsudera and S. Yokoyama, *Tetrahedron Lett.*, 1998, **39**, 7323 and references cited therein.
- 7 For examples of related cyclisations see: Y. Özlü, D. E. Cladingboel and P. J. Parsons, *Tetrahedron*, 1994, **50**, 2183.
- 8 Yu. I. El'natanov and R. G. Kostyanovskii, *Izv. Akad. Nauk SSSR, Ser. Khim.*, 1988, 382 (*Chem. Abtsr.*, 1989, **110**, 23303).
- 9 N. L. Zaichenko, I. I. Chervin, V. N. Voznesenskii, Yu. I. El'natanov and R. G. Kostyanovskii, *Izv. Akad. Nauk SSSR, Ser. Khim.*, 1988, 779 (*Chem. Abtsr.*, 1989, **110**, 22952).
- 10 A. B. Smith III, S. J. Branca, M. A. Guaciaro, P. M. Wovkulich and A. Korn, *Org. Synth.*, 1983, **61**, 65.
- 11 Y. Iwabuchi, M. Nakatani, N. Yokoyama and S. Hatakeyama, *J. Am. Chem. Soc.*, 1999, **121**, 10219 and references cited therein.
- 12 A. J. Pope, M. McVey, K. Fantom and K. J. Moore, *J. Biol. Chem.*, 1998, **273**, 31702 and references cited therein.